

## Cell adhesion and urothelial bladder cancer

Bryan, Richard T

DOI:

[10.1098/rstb.2014.0042](https://doi.org/10.1098/rstb.2014.0042)

License:

None: All rights reserved

Document Version

Peer reviewed version

Citation for published version (Harvard):

Bryan, RT 2015, 'Cell adhesion and urothelial bladder cancer: the role of cadherin switching and related phenomena', *Royal Society of London. Proceedings B. Biological Sciences*, vol. 370, no. 1661, 20140042. <https://doi.org/10.1098/rstb.2014.0042>

[Link to publication on Research at Birmingham portal](#)

### Publisher Rights Statement:

Final published version: Bryan, Richard T. "Cell adhesion and urothelial bladder cancer: the role of cadherin switching and related phenomena." *Philosophical Transactions of the Royal Society of London B: Biological Sciences* 370.1661 (2015): 20140042. <http://dx.doi.org/10.1098/rstb.2014.0042>

Checked October 2015

### General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

### Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact [UBIRA@lists.bham.ac.uk](mailto:UBIRA@lists.bham.ac.uk) providing details and we will remove access to the work immediately and investigate.

1    **CELL ADHESION AND UROTHELIAL BLADDER CANCER:**  
2    **THE ROLE OF CADHERIN SWITCHING AND RELATED PHENOMENA**

3  
4    **Richard T Bryan MBChB PhD MRCS**  
5    School of Cancer Sciences, College of Medical and Dental Sciences,  
6    University of Birmingham, UK

7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17    **Correspondence to:**    Dr Richard T Bryan, School of Cancer Sciences, University of Birmingham,  
18                                    Edgbaston, Birmingham B15 2TT, UK.  
19                                    r.t.bryan@bham.ac.uk, +44 121 414 7870.

20    **Running head:**            Cadherin cell adhesion in bladder cancer

21    **Keywords:**                Bladder cancer, cadherin, catenin, cell adhesion, stem cells, EpCAM

22    **Word count:**              4621 (Manuscript) + 157 (Abstract) + 4 Figures

23

## ABSTRACT

Cadherins are mediators of cell-cell adhesion in epithelial tissues. E-cadherin is a known tumour suppressor and plays a central role in suppressing the invasive phenotype of cancer cells. However, the abnormal expression of N- and P-cadherin ("cadherin switching") has been shown to promote a more invasive and malignant phenotype of cancer, with P-cadherin possibly acting as a key mediator of invasion and metastasis in bladder cancer. Cadherins are also implicated in numerous signalling events related to embryonic development, tissue morphogenesis, and homeostasis. It is these wide-ranging effects and the serious implications of cadherin switching that make the cadherin cell adhesion molecules and their related pathways strong candidate targets for the inhibition of cancer progression, including bladder cancer. This review will focus on cadherin switching in the context of bladder cancer and in particular the switch to P-cadherin expression, and will discuss other related molecules and phenomena, including EpCAM and the development of the cancer stem cell phenotype.

## MEDIA SUMMARY

Cadherins are mediators of cell-cell adhesion in epithelial tissues. E-cadherin is a tumour suppressor and plays a central role in suppressing the invasive phenotype of cancer cells. However, the abnormal expression of other cadherins ("cadherin switching") has been shown to promote a more invasive and malignant phenotype of cancer. Cadherins are also implicated in numerous signalling events related to embryonic development, tissue morphogenesis, and homeostasis. It is these wide-ranging effects and the serious implications of cadherin switching that make the cadherin cell adhesion molecules and related pathways attractive targets for the inhibition of cancer progression, including bladder cancer.

## BLADDER CANCER

### Introduction

Urothelial bladder cancer (UBC) is the fifth most common cancer in Western society, with a global incidence of over 356,000 and a prevalence estimated at 2.7million [1;2]. The burden of the disease is predicted to increase significantly in the foreseeable future as a result of population aging and the increasing world population, together with the progression of the tobacco epidemic and increasing exposure to occupational carcinogens in developing countries [2]. In the UK there are approximately 10,200 new cases and 5,000 deaths attributed to bladder cancer per year [3]. In Western populations over 90% of bladder cancers are transitional cell carcinomas of urothelial origin (urothelial cancers, UCs), and at presentation 75-85% will be non-muscle-invasive tumours (NMIBC, stages Ta/T1/Tis), with the remainder being muscle-invasive (MIBC, stages T2-4) [1;4-6].

NMIBC is a heterogeneous disease typified by a high rate of recurrence (15-61% at one year, depending upon risk category [7]) and so long-term, even lifelong, surveillance with outpatient flexible cystoscopy is the mainstay of subsequent management [6;8]. Progression to MIBC is also a concern for high-risk NMIBC patients, occurring in up to 17% of patients at one year [7]. However, the overall prognosis is good with 65-85% of patients surviving for 5 years or more [5].

Progression to (or presentation with) MIBC represents the critical step in the disease course, necessitating more radical therapies and carrying a 5-year survival rate of only 25-50% [5;9]. For curative intent, patients who present with or progress to MIBC are treated by radiotherapy [6;10], chemoradiotherapy [11], radical cystectomy, or neoadjuvant chemotherapy followed by radical cystectomy [6;9;10].

The cumulative cost of treating UBC exceeds all other forms of human cancer, the majority of which is attributable to the long-term treatment and surveillance of NMIBC [12-14]. Despite this, there is only modest research funding for UBC compared to other malignancies [15], and as a result there has been a lack of scientific advancement in the field [15-17], with no major new drugs approved for UBC in over 10-years [17;18].

Cadherins are mediators of cell-cell adhesion in epithelial tissues [19;20]. We have previously demonstrated that the abnormal expression of P-cadherin (an example of “cadherin switching”) is associated with an invasive and

aggressive phenotype of UBC [21], and have hypothesized that P-cadherin may act as a key effector of muscle-invasion [22]. The cadherins are involved in a number of important phenomena related to cancer progression, including epithelial-to-mesenchymal transition (EMT) and the development of a cancer stem cell phenotype [22;23]. It is these wide-ranging effects and the serious implications of cadherin switching that make the cadherins and their related pathways strong candidate targets for the inhibition of cancer progression, including UBC. This review will focus on cadherin-based cell adhesion in the context of UBC and the switch to P-cadherin expression, and will discuss other related molecules and phenomena, including EpCAM and the development of the cancer stem cell phenotype.

## **METHODS**

Our group has been working in the field of cadherin biology for a number of years [24;25], and we regularly review the literature on these molecules and their associated pathways [22]. Specifically, this review was written utilising papers obtained following *PubMed* searches and with the following structure: bladder cancer background, epidemiology and molecular pathogenesis; cadherin background and biology; cadherins in epithelial malignancies, cadherin switching, and cadherins in bladder cancer. The background to cadherins and cadherin biology presented here has been derived from key papers by workers who initially characterised and described these molecules, and then who subsequently investigated cadherin expression and function in various epithelial malignancies and model systems. We updated the field for cadherin switching to describe this process in the context of malignancy and related phenomena (eg. epithelial-to-mesenchymal transition, cell migration, metastasis, cancer stem cells, EpCAM signalling), utilising papers written by significant workers in this field. The data, findings and information contained within these publications were then assimilated to create a review of cadherin switching in bladder cancer and including some of our own interpretations.

## **MOLECULAR PATHWAYS TO NON-MUSCLE-INVASIVE & MUSCLE-INVASIVE BLADDER CANCER**

Different approaches have been taken to describe the molecular alterations involved in bladder tumorigenesis [26-31]. We have previously described such pathways based upon the six “hallmarks of cancer” described by Hanahan

and Weinberg in 2000 [32-35]. In 2011 Hanahan and Weinberg updated their original landmark review, describing genome instability and inflammation as underlying these hallmark changes, and proposed “reprogramming of energy metabolism” and “evading immune destruction” as two emerging hallmarks with potential for generality [35]. In addition, they described that tumors exhibit another dimension of complexity by containing a repertoire of recruited, ostensibly normal cells that contribute to the acquisition of hallmark traits by creating the “tumor microenvironment” [35], and our own research has demonstrated the apparent importance of the immunological milieu of the bladder tumour microenvironment [RT Bryan et al - unpublished data]. In their 2011 update, Hanahan and Weinberg also introduced the concept of “cancer stem cells” [35], a concept that has existed for a number of years in haematopoietic malignancies [36;37]. Cancer stem cells (CSCs) are a subset of tumor cells that have the ability to self-renew and to generate all of the heterogeneous cells that comprise a tumor (properties that are analogous to a stem cell, the original cell of an organ and responsible for organogenesis and organ maintenance) [23;36;38-40]. In the setting of UBC, CSCs appear to play a role in a subset of tumors, but their true significance is yet to be clarified [23].

Other authors have reviewed the field of UBC molecular pathogenesis in detail [26-31], and there has been general consensus on a divergent pathway for the development of Ta/T1 disease and Tis/T2+ disease [29;41-46]. However, Dancik et al recently identified a cell of origin gene signature for basal cells and umbrella cells of the urothelium [47]. By utilising this cell of origin signature in UBCs from 874 patients, it appeared that NMIBCs and MIBCs developed from distinct progenitor cells [47], possibly shifting our understanding of urothelial carcinogenesis away from the classical two pathway model. Further detailed genomic and epigenomic studies of both MIBCs and NMIBCs are thus required to clarify our understanding of the pathogenesis of these tumours [48].

Although a detailed examination of these pathways is beyond the scope of this review, this is a rapidly changing field and new developments appear frequently with the advent of high-throughput experimental platforms including “deep sequencing” [49], proteomics [50-52] and metabolomics [53]. Most recently, The Cancer Genome Atlas (TCGA) Research Network undertook the comprehensive molecular characterization of 131 MIBCs [49]. With regard to somatic DNA mutations, a notable finding was the significant enrichment of non-silent mutations in chromatin regulatory genes compared to other epithelial cancers studied: 76% of the tumours (MIBCs) had an inactivating

mutation in one or more of these genes, and 41% had at least two such mutations [49]. *TP53* mutations were also common (49%), as were amplification and overexpression of *MDM2*, suggesting that TP53 function was inactivated in 76% of tumours [49]. There were a large number of previously undescribed mutations, and viral DNAs and transcripts were also identified [49]. RNA-seq data identified 4 tumour clusters and pathway analysis demonstrated three frequently dysregulated pathways [49]: cell-cycle regulation (altered in 93% of cases); kinase and phosphatidylinositol-3-OH kinase (PI(3)K) signaling (72%); and chromatin remodelling (89%). A number of the genomic alterations identified are theoretically amenable to therapeutic targeting [49], and such new therapeutics are desperately needed for UBC [17;18;54].

Choi et al also utilised whole genome mRNA expression profiling to cluster MIBCs into 3 distinct groups, based upon the established molecular subtypes of breast cancer [55]: basal MIBCs shared biomarkers with basal breast cancers and were characterized by p63 activation, squamous differentiation, and more aggressive disease; luminal MIBCs contained features of active PPAR $\gamma$  and oestrogen receptor transcription and were enriched with activating *FGFR3* mutations and potential *FGFR* inhibitor sensitivity; p53-like MIBCs were consistently resistant to a number of chemotherapeutics, including cisplatin; and all chemoresistant tumours adopted a p53-like phenotype after therapy [55]. These findings have important implications for the clinical management of MIBC: they include not only prognostic information, but also suggestions for subtype-directed targeted therapy and potential to predict response to cisplatin-based chemotherapy (although further work is needed to elucidate other biomarkers of resistance) [56]. It is, however, disappointing that NMIBCs were not analysed in the same way by either the TCGA Research Network or Choi et al [48], especially as these tumours represent the vast majority (>75%) of bladder cancer patients [57;58].

## **CADHERINS**

The classical cadherins are calcium-dependent transmembrane glycoproteins found at the adherens junction and are mediators of cell-cell adhesion in epithelial tissues [19;20]. E-cadherin is a tumour suppressor, playing a central role in suppressing the invasive phenotype of UBC cells [59]. The abnormal expression of other “classical” cadherins (P- and N-cadherin) has been shown to promote a more invasive and malignant phenotype of UBC [24], possibly acting

145 as key mediators of invasion and metastasis. With such a large difference in UBC outcomes between early stage  
146 disease (stage Ta) versus MIBC (stages T2+) it is reasonable to assume that cell adhesion molecules, and in particular  
147 cadherins, play a fundamental role in the spread of bladder tumours, initially from the urothelium into the lamina  
148 propria (through the basement membrane) and subsequently into the detrusor muscle [22]. Therefore, the classical  
149 cadherins and their related molecular pathways represent attractive therapeutic targets for the inhibition of  
150 progression in bladder cancer patients [19;59-61].

151 Cadherins comprise of extracellular (EC1-5), transmembranous, and cytoplasmic domains, with the cytoplasmic  
152 domain anchored to the cell cytoskeleton by catenin family members ( $\alpha$ -,  $\beta$ -,  $\gamma$ -catenin and p120) [19;61-65]. P21-  
153 activated kinase 5 (PAK5) also appears to associate with  $\beta$ -catenin and p120 to stabilise the adherens junction in  
154 order to maintain normal cell-cell adhesion [66]. Traditionally, cell-cell adhesion is described as being achieved by  
155 the symmetric interactions of the first extracellular domains (EC1) of cadherins on neighbouring cells (trans-  
156 interaction) [64;67]; cadherins on the same cell also interact with each other (cis-interaction) through the EC1  
157 domain of one and the EC2 domain of the other [64;67;68]. More recently, it has been described that optimal cell-  
158 cell adhesion (50-70pN) is achieved by all 5 EC domains of E-cadherin, and with a cell-cell separation of 5-11nm [65].  
159 See **Figure 1**. E-, P- and N-cadherin were the first cadherins identified, and can all mediate cell-cell adhesion in this  
160 fashion [63;69]:

- 161 • E-cadherin (CDH1, 120kDa): the main mediator of cell-cell adhesion in epithelial tissues and expressed by  
162 most normal epithelial cells [19;61;62;69-71].
- 163 • N-cadherin (CDH2, 130kDa): expressed by neural, endothelial, and muscle cells, but not normally by  
164 epithelial cells [62;69].
- 165 • P-cadherin (CDH3, 118kDa): normally only weakly expressed in the basal layers of stratified epithelia such as  
166 oesophagus, bronchus and bladder [24;69;71].

167 Epithelial malignancies, including bladder cancer, typically show loss of E-cadherin expression as grade and stage  
168 progress, and this is often accompanied by increased expression of N- or P-cadherin. This phenomenon is described  
169 as “cadherin switching” [19;61;69;71-73], illustrated in the bladder cancer setting in **Figure 2**. Excellent reviews of



170 the field have been published recently [74;75], and we have previously reviewed this field for bladder cancer [22];  
171 we provide an overview below.

172

## 173 CADHERIN SWITCHING

174 Cadherin switching (CS) is a hallmark of epithelial-to-mesenchymal transition (EMT) [76], the process by which  
175 epithelial cells lose their characteristic polarity, disassemble cell junctions, and become more migratory as a  
176 precursor to invasion and metastasis (they acquire properties analogous to mesenchymal cells) [19;25;61;77-82]. In  
177 this setting, CS typically describes a process where the normal expression of E-cadherin is replaced by the abnormal  
178 expression of N-cadherin, or where N-cadherin expression is increased and E-cadherin levels remain unchanged  
179 [19;61;76]. CS appears to play a role late in many malignancies (including breast, prostate, pancreas, ovarian,  
180 bladder and melanoma), resulting in a more invasive and malignant phenotype of disease with a worse outcome  
181 [19;24;61;74-76;83-89]. The regulation of CS is yet to be fully elucidated, but most likely involves transcriptional and  
182 post-transcriptional events, possibly influenced by cytokines or growth factors [19;61]. Recently, Slug (*SNAI2*, a  
183 member of the Snail family of zinc-finger transcription factors) has been identified to play a critical role in EMT by  
184 control of the E-cadherin to N-cadherin switch in UBC [90].

185 In UBC, ourselves and others have described CS, demonstrating increased expression of both P- and N-cadherin in  
186 late stage high-grade disease (**Figure 2**) [24;69;89;91;92]. We studied 153 bladder tumours and utilised a variety of  
187 cell lines and functional in vitro models [24]: increased membranous P-cadherin expression was observed in almost  
188 half of all MIBCs and almost 40% of grade 3 UBCs, accompanied by significantly reduced expression of E-cadherin  
189 [24]. Increased P-cadherin expression was associated with worse bladder cancer-specific survival, and P-cadherin  
190 status was an independent prognostic factor (alongside grade and stage) [24]. Functional in vitro experiments  
191 showed that altering the balance of E- and P-cadherin in favour of P-cadherin expression enhanced anchorage-  
192 independent growth, and that P-cadherin alone was unable to mediate normal cell-cell adhesion [24]. We concluded  
193 that P-cadherin expression promoted a more malignant and invasive phenotype of bladder cancer (even in the  
194 presence of E-cadherin), and appeared to have a novel role late in the disease process [24].

195 Mandeville et al also demonstrated similar findings [92]. In their in vitro studies, utilising P-cadherin transfection and  
196 knockdown, they demonstrated that P-cadherin induced a significant increase in migratory capacity (although with  
197 no accompanying change in invasive potential) [92]. The authors suggested that P-cadherin may have a role in  
198 regulating the migration of basal cells to the intermediate cell layer in normal urothelium, as well as a role in  
199 neoplastic progression [92]. More recently, Wang et al have demonstrated similar findings [89].

200 Ourselves and others have postulated that a subgroup of aggressive P-cadherin-expressing tumours may be derived  
201 from the normally weakly P-cadherin-expressing basal layer of the urothelium [22]. In support of this hypothesis,  
202 Van Batavia et al recently demonstrated that papillary and CIS lesions were derived from different urothelial  
203 populations, with intermediate cells contributing to non-invasive papillary lesions and basal cells representing the  
204 origin of CIS (which ultimately leads to MIBC) [93]. These findings support a model in which the heterogeneity  
205 observed in bladder cancers is determined both by genetic changes and the cell lineage from which the tumour  
206 originates [93].

207 However, despite P-cadherin expression being associated with a more aggressive phenotype in many cancers, such  
208 behaviour is not ubiquitous and is context dependent [75]. For example, in malignant melanoma, which commonly  
209 demonstrates a cadherin switch to N-cadherin expression [22], P-cadherin promotes adhesion and inhibits invasion  
210 in a similar fashion to E-cadherin [75], and E-cadherin negative breast cancer cells show many similarities when  
211 subsequently transfected with E- or P-cadherin [74;94]. Ribeiro et al investigated these phenomena in detail in a  
212 breast cancer model, demonstrating that P-cadherin co-localizes with E-cadherin, and promotes cell invasion by  
213 disrupting E-cadherin/catenin interactions [95]. E- and P-cadherin co-expressing tumour cells showed enhanced in  
214 vivo tumour growth compared with those expressing only E- or only P-cadherin, and co-expression of E- and P-  
215 cadherin in breast tumours correlated with high-grade biologically aggressive tumours accompanied by poor patient  
216 survival [95]. It is therefore feasible that P-cadherin only promotes invasion in tissues that endogenously express E-  
217 cadherin [74], with heterodimerisation between E- and P-cadherin disrupting the formation of functional cadherin-  
218 catenin complexes [75].

219 It is likely that the key mechanisms involved in P-cadherin's deregulation largely occur in the promoter region of  
220 *CDH3* and not by structural alterations of its coding sequences [74]: in 2005, Paredes et al demonstrated

hypomethylation of the *CDH3* gene promoter correlated with P-cadherin overexpression in breast cancer [74;96], and other workers have described this phenomenon in pancreatic [74;97] and colorectal cancers [74;98]. Our own data suggest differential *CDH3* promoter methylation between bladder cancer cell lines and tumours, and normal urothelium [RT Bryan - unpublished data]. Furthermore, the balance of E- and P-cadherin expression impacts the overall genetic programme [74], altering the expression of genes involved in signal transduction and growth factors, cell cycle, cell adhesion and the extracellular matrix, cytokines and inflammation [74;94]. In addition, P-cadherin can provoke the secretion of pro-invasive factors such as the matrix metalloproteinases MMP1 and MMP2 [74;75;99]. The role of p120 also appears important, with P-cadherin probably interfering with the normal binding of p120 to E-cadherin at the adherens junction [74;100]. In a pancreatic cancer model, accumulation of p120 in the cytoplasm (and not bound to E-cadherin at the membrane) appeared to induce the increased cell migration seen following P-cadherin expression via the Rho GTPases, Rac1 and Cdc42 [74;101]. P-cadherin-induced increase in Rac1 and Cdc42 activity (mediated via p120) has also been observed in ovarian cancer [74;102]. Specifically, insulin-like growth factor 1 receptor (IGF1R) can seemingly form a complex with P-cadherin, resulting in the tyrosine phosphorylation and activation of cytoplasmic p120 to promote invasion [75;102;103]; this pathway appears specific to P-cadherin and not the other classical cadherins [75;103].

Taken together, all of the data above emphasise that P-cadherin represents a very attractive target for novel anti-cancer therapeutics [74], and phase I trials of a P-cadherin inhibitor (PF-03732010, a human monoclonal antibody against P-cadherin) have been undertaken [104], although its development now seems to have stalled.

## **CADHERINS AND CANCER STEM CELLS**

Although solid tumours can be reduced in size or eradicated by chemotherapy, radiotherapy or surgery (alone or in combinations), disease relapse or progression often occurs [105;106]. Such relapse or progression may be explained by the persistence of residual tumour-initiating cells and tumour-maintaining cells, and such cells have been reported in a variety of malignancies (breast, brain, prostate, lung, pancreas, etc) since they were first identified in leukaemia [79;105;107]. Such “cancer stem cells” (CSCs) theoretically have the ability to self-renew and to generate

246 the heterogeneous cells that comprise a tumour [105-110], and thus need to be eradicated to provide long-term  
247 disease-free survival (although it appears that CSCs are more resistant to conventional therapies) [108;110-112].  
248 CSCs may either develop following genetic or epigenetic events in normal stem cells or from differentiated tumour  
249 cells that develop the capability for unlimited growth [23;82]. Cellular markers of “stemness” are still under debate,  
250 but include CD44, CD24, CD133 and EpCAM [82]: in breast, prostate and oral squamous carcinomas, CSCs are likely  
251 identified as CD44<sup>+</sup>/CD24<sup>-</sup>, whereas CD133 appears to be a CSC marker in gliomas and in colon and pancreatic  
252 carcinomas [82].

253 In a previous review we suggested that the evidence supports the CSC paradigm for UBC, as in other epithelial  
254 malignancies [23]. As discussed above, in normal urothelium P-cadherin is only expressed in the basal cell layer (the  
255 assumed urothelial stem cell niche) and in a subset of more aggressive UBCs [21-23;92;113]. It is therefore tempting  
256 to assume that P-cadherin is a marker of urothelial stem cells and UBC CSCs. Although E-cadherin intercellular  
257 adhesion is considered important for the survival of human embryonic stem cells (hESCs) and induced pluripotent  
258 stem cells (iPSCs) [82], Kolle et al recently identified *CDH3* (P-cadherin) and *TACSTD1* (EpCAM) as genes encoding  
259 hESC markers (antibodies for EpCAM were also able to enrich for pluripotent hESCs) [114]. Vieira et al have also  
260 demonstrated that P-cadherin mediates stem cell properties in basal-like breast cancer [115]. P-cadherin therefore  
261 appears promising as a potential marker of CSCs in UBC, and similar work is required to confirm these findings in  
262 UBC [23]. The fact that *CDH3* (P-cadherin) did not appear in Dancik et al’s cell of origin signature described earlier is  
263 somewhat surprising since it is normally expressed by basal urothelial cells and in a subset of aggressive UBCs that  
264 may also harbour CSCs; however, as described above, P-cadherin’s deregulation is most likely governed by  
265 epigenetic phenomena rather than structural alterations in its coding sequences [74]. Characterisation of the UBC  
266 epigenome/methylome may thus be required to elucidate P-cadherin’s role in these UBC subtypes.

267 It is highly feasible that treatment-resistant cells develop via other mechanisms and pathways, with CSCs being  
268 responsible only for a minority [105;116]. Heterogeneity within some tumours may result from selective pressure  
269 during tumorigenesis [35;112]. See **Figure 3**. It has been suggested that UBCs arise from more differentiated cells,  
270 and self-renewal capacity may be acquired secondarily by inactivation of *p53* and *RB1* function [105;116]. The  
271 tumour microenvironment may also play an important role [108], potentially inducing a transitory or reversible CSC-

272 like state [117]: although EMT may drive the development of CSCs [35], EMT itself is reversible with mesenchymal-  
273 to-epithelial transition (MET) favouring a cell's colonisation of distant sites to generate metastases [35]. Whether the  
274 CSC state reverses in a similar setting and fashion remains unknown, but such interactions highlight the importance  
275 of the tumour microenvironment for all cancer cells, not just CSCs [35].

276

## 277 CADHERINS AND EPCAM

278 EpCAM is a type-1 membrane protein that functions as a cell adhesion molecule [118]. It is overexpressed in many  
279 epithelial malignancies, including bladder CIS [119] and high grade and advanced stage UBCs [120]. The tumour-  
280 specific expression of EpCAM has led to its use for capturing circulating tumour cells by the FDA-approved  
281 *CELLSEARCH* system [121], and also for directing therapies to bladder tumours [122]. High tissue levels of EpCAM are  
282 associated with a poor prognosis in UBC [120]. However, the role of EpCAM remains elusive: both tumour  
283 suppressor and oncogenic properties have been reported. In 2009, Maetzel et al demonstrated that EpCAM could be  
284 sequentially cleaved to release extracellular and intracellular domains, 'EpEX' and 'EpICD', respectively [123]; EpICD  
285 diffuses into the nucleus and activates oncogenic signalling events by associating with FHL2,  $\beta$ -catenin and Lef-1  
286 [123;124]. See **Figure 4**.

287 In 2014, as part of our de novo urinary biomarker discovery programme [125], we demonstrated that elevated  
288 urinary EpCAM was observed in patients with grade 3 NMIBCs and MIBCs [51;52]. EpCAM was a significant  
289 independent prognostic factor for UBC-specific survival, with elevated urinary levels resulting in an increased risk of  
290 dying from bladder cancer (hazard ratio 1.76). The predominant form of EpCAM in the urine was a soluble and stable  
291 form comprised of the entire extracellular domain, and not the intact protein [52]. Our data therefore suggested  
292 that the cleavage of EpCAM into EpEX and EpICD could also occur in UBC [52;123], and further evidence supports  
293 this: Ralhan *et al* recently demonstrated that 9 out of 10 cases of UBC were positive for EpICD [126]. However, our  
294 work demonstrated that the extracellular domain of EpCAM was released by cleavage immediately adjacent to the  
295 cell membrane [52]; the exact location of cleavage was not described by Maetzel et al [123], but the protease

involved (TACE or ADAM 17) usually cleaves membrane proteins 10-15 residues away from the membrane surface [127], suggesting atypical cleavage or an alternative mechanism of extracellular domain release in UBC [52].

Notably, there are important relationships between EpCAM and classical cadherins, although this relationship appears to be tissue- and tumour-specific [128]. In 1997, Litvinov et al suggested that EpCAM has a role in the development of a proliferative and malignant phenotype of epithelial cell [129]: increasing the expression of EpCAM in cadherin-positive cells led to the gradual abrogation of adherens junctions [129]. Although EpCAM had no influence on the total amount of cellular cadherin, it affected the interaction of the cadherins with the cytoskeleton and, as cadherin-mediated cell-cell adhesion diminished, EpCAM-mediated intercellular connections predominated [129]. In a murine fibroblast model, Winter et al subsequently demonstrated that this may occur by disruption of the link between  $\alpha$ -catenin and F-actin, probably by EpCAM's disruption of the actin cytoskeleton or possibly via p120 [130]. In later work on human breast epithelial cells, the same authors demonstrated that EpCAM cross-signaling with N-cadherin resulted in the abrogation of cadherin adhesion complexes, mediated by PI(3)K [131]. In breast cancer cell lines, Martowicz et al showed that epithelial cells need EpCAM to promote growth and invasion, yet mesenchymal tumour cells are independent of EpCAM for invasion and progression [132]; the same authors also demonstrated that overexpression of EpCAM in human mammary epithelial cells led to a more proliferative phenotype and downregulation of E-cadherin [133].

Conversely, in a zebrafish model, Slanchev et al demonstrated that EpCAM was indispensable for skin epithelial integrity, and that *epcam* mutant embryos displayed reduced levels of membranous E-cadherin [134]. Guerra et al also postulated an important role for EpCAM in the maintenance of normal intestinal architecture and function in congenital tufting enteropathy, utilising an *mTrop1/Epcam* knockout mouse model of the disease [135]. Other model systems have also demonstrated a direct association between loss of EpCAM expression and loss of cadherin-mediated adhesion [136].

Seemingly, EpCAM has dual functions in normal and cancerous cells with regard to cadherin regulation, cell-cell adhesion and epithelial integrity: EpCAM may be essential for normal epithelial tissue integrity and cell-cell adhesion, but there also appears to be a role for EpCAM in the disruption of normal cell-cell adhesion to initiate EMT, with the subsequent transformed cells acting independently of EpCAM signaling for invasion and progression.

322 Interestingly, Zeb1 (a known transcription factor inducing EMT) represses both E-cadherin and EpCAM by binding to  
323 the EpCAM promoter [137], yet the expression of E-cadherin and EpCAM is related to a stem cell-like phenotype  
324 [138;139]; in basal-like breast cancer EpCAM and P-cadherin both appear to be associated with the CSC phenotype  
325 [115]. As described for the hallmarks of cancer [34], the timing and ordering of these events appears to differ  
326 between normal and tumorous tissues, between different tissue and tumour types, and most likely within the same  
327 tumour. It is feasible that during EMT in some malignancies, EpCAM may stimulate the dissolution of E-  
328 cadherin/catenin complexes and so permit P- and N-cadherin complexes to predominate (cadherin switching) and  $\beta$ -  
329 catenin-mediated oncogene transcription to be upregulated; yet in other tumour types, EpCAM and E-cadherin may  
330 be downregulated in parallel, with EMT being driven by alternate pathways. Conversely, EpCAM may stabilise E-  
331 cadherin/catenin complexes in some tumours, possibly providing a "stable" and less chaotic cellular milieu  
332 unaffected by EMT, in which the development of a CSC phenotype can be "nurtured" by alternative pathways (as  
333 described above, EpCAM is a cell surface marker of hESCs, and can be used to isolate a pluripotent subpopulation  
334 from hESC culture [114]). If the latter model is correct, then the corollary would potentially be the normalisation of  
335  $\beta$ -catenin-mediated transcription in CSCs; evidence to date in other malignancies suggests that this is not the case  
336 [140-142]. However, these are dynamic processes, and even within the same tumour all of these proposed  
337 phenomena may be unfolding simultaneously; in the future, single cell genomics may resolve these issues [143;144].  
338 It is important to note that CSC-like treatment-resistant disease may develop via alternate pathways (Figure 3), and  
339 there is likely to be considerable plasticity [142], with cells reverting to a less aggressive state by mesenchymal-to-  
340 epithelial transition (MET) or by the reversal of the CSC phenotype. Furthermore, the influence of EpCAM on P-  
341 cadherin is yet to be elucidated. Our current research is attempting to resolve some of these mechanisms.

## DISCUSSION & CONCLUSION

P-cadherin seemingly has a number of fundamental roles in bladder cancer and other malignancies, including mediating the development of CSCs and EMT, both of which lead to more aggressive disease and worse survival. The mechanisms of these phenomena have been well-described in other malignancies, but remain to be elucidated in UBC. Although we have assumed some crossover of P-cadherin's function between tumour and tissue types, we know that many of P-cadherin's actions are tumour- and tissue-specific. Therefore, such findings from other malignancies need to be reproduced in UBC if we are to genuinely understand P-cadherin's role in this setting. However, given the genomic characterizations of MIBC described above [47;49;55], it is unlikely that P-cadherin represents a "driver" of urothelial carcinogenesis [145]; P-cadherin is more likely to represent an important downstream effector of such driver mutations, with multiple influences on important pathways and phenomena that determine outcomes in advanced disease (eg. EMT, CSCs), probably mediated by PI(3)K [49]. Moreover, it appears that P-cadherin plays a fundamental role in the cell surface and cell adhesion phenomena that permit tumour cells to migrate and invade, and possibly to metastasize.

In conclusion, P-cadherin represents a highly attractive therapeutic target, alongside N-cadherin [146-148]. However, given P-cadherin's complex interactions described above (and undoubtedly many yet to be discovered), P-cadherin inhibition may have far more wide-reaching effects than those directly related to tumour invasion and progression. The difficulties of taking an anti-P-cadherin agent through clinical trials and into clinical use should therefore not be underestimated. Furthermore, the association of classical cadherins with EpCAM is particularly fascinating and requires further elucidation in UBC, and our work in this area is ongoing.



- (1) van Rhijn BW, Burger M, Lotan Y, Solsona E, Stief CG, Sylvester RJ, Witjes JA, Zlotta AR. Recurrence and Progression of Disease in Non-Muscle-Invasive Bladder Cancer: From Epidemiology to Treatment Strategy. *Eur Urol* 2009.
- (2) Ploeg M, Aben KK, Kiemeny LA. The present and future burden of urinary bladder cancer in the world. *World J Urol* 2009; 27:289-293.
- (3) CRUK. CancerStats Key Facts - Bladder Cancer. 2009.  
Ref Type: Report
- (4) Lorusso V, Silvestris N. Systemic chemotherapy for patients with advanced and metastatic bladder cancer: current status and future directions. *Ann Oncol* 2005; 16 Suppl 4:iv85-iv89.
- (5) Wallace DM, Bryan RT, Dunn JA, Begum G, Bathers S. Delay and survival in bladder cancer. *BJU Int* 2002; 89:868-878.
- (6) Kaufman DS, Shipley WU, Feldman AS. Bladder cancer. *Lancet* 2009; 374:239-249.
- (7) Sylvester RJ, van der Meijden AP, Oosterlinck W, Witjes JA, Bouffieux C, Denis L, Newling DW, Kurth K. Predicting recurrence and progression in individual patients with stage Ta T1 bladder cancer using EORTC risk tables: a combined analysis of 2596 patients from seven EORTC trials. *Eur Urol* 2006; 49:466-5.
- (8) Babjuk M, Burger M, Zigeuner R, Shariat SF, van Rhijn BW, Comperat E, Sylvester RJ, Kaasinen E, Bohle A, Palou RJ, Roupret M. EAU Guidelines on Non-Muscle-invasive Urothelial Carcinoma of the Bladder: Update 2013. *Eur Urol* 2013.
- (9) Neoadjuvant chemotherapy in invasive bladder cancer: update of a systematic review and meta-analysis of individual patient data advanced bladder cancer (ABC) meta-analysis collaboration *Eur Urol* 2005; 48:202-205.
- (10) Stenzl A, Cowan NC, De SM, Kuczyk MA, Merseburger AS, Ribal MJ, Sherif A, Witjes JA. Treatment of muscle-invasive and metastatic bladder cancer: update of the EAU guidelines. *Eur Urol* 2011; 59:1009-1018.
- (11) James ND, Hussain SA, Hall E, Jenkins P, Tremlett J, Rawlings C, Crundwell M, Sizer B, Sreenivasan T, Hendron C, Lewis R, Waters R, Huddart RA. Radiotherapy with or without chemotherapy in muscle-invasive bladder cancer. *N Engl J Med* 2012; 366:1477-1488.
- (12) Botteman MF, Pashos CL, Redaelli A, Laskin B, Hauser R. The health economics of bladder cancer: a comprehensive review of the published literature. *Pharmacoeconomics* 2003; 21:1315-1330.
- (13) Riley GF, Potosky AL, Lubitz JD, Kessler LG. Medicare payments from diagnosis to death for elderly cancer patients by stage at diagnosis. *Med Care* 1995; 33:828-841.
- (14) Simons MP, O'Donnell MA, Griffith TS. Role of neutrophils in BCG immunotherapy for bladder cancer. *Urol Oncol* 2008; 26:341-345.
- (15) Lotan Y, Kamat AM, Porter MP, Robinson VL, Shore N, Jewett M, Schelhammer PF, deVere WR, Quale D, Lee CT. Key concerns about the current state of bladder cancer: a position paper from the Bladder Cancer Think Tank, the Bladder Cancer Advocacy Network, and the Society of Urologic Oncology. *Cancer* 2009; 115:4096-4103.

402 (16) Kaplan AL, Litwin MS, Chamie K. The future of bladder cancer care in the USA. *Nat Rev Urol* 2014; 11:59-62.

403 (17) Bryan RT, Kirby R, O'Brien T, Mostafid H. So Much Cost, Such Little Progress. *Eur Urol* 2014.

404 (18) Bryan RT, James ND. Bladder cancer: time for a rethink? *Oncology (Williston Park)* 2011; 25:965, 968.

405 (19) Cavallaro U, Schaffhauser B, Christofori G. Cadherins and the tumour progression: is it all in a switch? *Cancer*  
406 *Lett* 2002; 176:123-128.

407 (20) Takeichi M. Morphogenetic roles of classic cadherins. *Curr Opin Cell Biol* 1995; 7:619-627.

408 (21) Bryan RT, Atherfold PA, Yeo Y, Jones LJ, Harrison RF, Wallace DM, Jankowski JA. Cadherin switching dictates  
409 the biology of transitional cell carcinoma of the bladder: ex vivo and in vitro studies. *J Pathol* 2008; 215:184-  
410 194.

411 (22) Bryan RT, Tselepis C. Cadherin switching and bladder cancer. *J Urol* 2010; 184:423-431.

412 (23) Bryan RT. Bladder cancer and cancer stem cells: basic science and implications for therapy.  
413 *ScientificWorldJournal* 2011; 11:1187-1194.

414 (24) Bryan RT, Atherfold PA, Yeo Y, Jones LJ, Harrison RF, Wallace DM, Jankowski JA. Cadherin switching dictates  
415 the biology of transitional cell carcinoma of the bladder: ex vivo and in vitro studies. *J Pathol* 2008; 215:184-  
416 194.

417 (25) Hardy RG, Tselepis C, Hoyland J, Wallis Y, Pretlow TP, Talbot I, Sanders DS, Matthews G, Morton D, Jankowski  
418 JA. Aberrant P-cadherin expression is an early event in hyperplastic and dysplastic transformation in the  
419 colon. *Gut* 2002; 50:513-519.

420 (26) Birkhahn M, Mitra AP, Cote RJ. Molecular markers for bladder cancer: the road to a multimarker approach.  
421 *Expert Rev Anticancer Ther* 2007; 7:1717-1727.

422 (27) Castillo-Martin M, Domingo-Domenech J, Karni-Schmidt O, Matos T, Cordon-Cardo C. Molecular pathways of  
423 urothelial development and bladder tumorigenesis. *Urol Oncol* 2010; 28:401-408.

424 (28) Cheng L, Zhang S, MacLennan GT, Williamson SR, Lopez-Beltran A, Montironi R. Bladder cancer: translating  
425 molecular genetic insights into clinical practice. *Hum Pathol* 2011; 42:455-481.

426 (29) Knowles MA. Molecular pathogenesis of bladder cancer. *Int J Clin Oncol* 2008; 13:287-297.

427 (30) Shariat SF, Karam JA, Lerner SP. Molecular markers in bladder cancer. *Curr Opin Urol* 2008; 18:1-8.

428 (31) Youssef RF, Mitra AP, Bartsch G, Jr., Jones PA, Skinner DG, Cote RJ. Molecular targets and targeted therapies  
429 in bladder cancer management. *World J Urol* 2009; 27:9-20.

430 (32) Bryan RT, Hussain SA, James ND, Jankowski JA, Wallace DM. Molecular pathways in bladder cancer: part 1.  
431 *BJU Int* 2005; 95:485-490.

432 (33) Bryan RT, Hussain SA, James ND, Jankowski JA, Wallace DM. Molecular pathways in bladder cancer: part 2.  
433 *BJU Int* 2005; 95:491-496.

434 (34) Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000; 100:57-70.

435 (35) Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011; 144:646-674.

436 (36) Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature* 2001;  
437 414:105-111.

438 (37) Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive  
439 hematopoietic cell. *Nat Med* 1997; 3:730-737.

440 (38) Brandt WD, Matsui W, Rosenberg JE, He X, Ling S, Schaeffer EM, Berman DM. Urothelial carcinoma: stem  
441 cells on the edge. *Cancer Metastasis Rev* 2009; 28:291-304.

442 (39) Gupta PB, Chaffer CL, Weinberg RA. Cancer stem cells: mirage or reality? *Nat Med* 2009; 15:1010-1012.

443 (40) Visvader JE, Lindeman GJ. Cancer stem cells in solid tumours: accumulating evidence and unresolved  
444 questions. *Nat Rev Cancer* 2008; 8:755-768.

445 (41) Goebell PJ, Knowles MA. Bladder cancer or bladder cancers? Genetically distinct malignant conditions of the  
446 urothelium. *Urol Oncol* 2010; 28:409-428.

447 (42) Knowles MA, Currie GA. Genetic alterations in bladder cancer. *Lancet* 1993; 342:1184.

448 (43) Knowles MA. What we could do now: molecular pathology of bladder cancer. *Mol Pathol* 2001; 54:215-221.

449 (44) Knowles MA. Molecular subtypes of bladder cancer: Jekyll and Hyde or chalk and cheese? *Carcinogenesis*  
450 2006; 27:361-373.

451 (45) Knowles MA. Bladder cancer subtypes defined by genomic alterations. *Scand J Urol Nephrol Suppl* 2008;116-  
452 130.

453 (46) Hurst CD, Platt FM, Knowles MA. Comprehensive Mutation Analysis of the TERT Promoter in Bladder Cancer  
454 and Detection of Mutations in Voided Urine. *Eur Urol* 2013.

455 (47) Dancik GM, Owens CR, Iczkowski KA, Theodorescu D. A cell of origin gene signature indicates human bladder  
456 cancer has distinct cellular progenitors. *Stem Cells* 2014; 32:974-982.

457 (48) Bryan RT, Kirby R, Mostafid H. Does the Nonurologic Scientific Community Understand Urothelial Bladder  
458 Cancer? *Eur Urol* 2014.

459 (49) Comprehensive molecular characterization of urothelial bladder carcinoma *Nature* 2014; 507:315-322.

460 (50) Bryan RT, Wei W, Shimwell NJ, Collins SI, Hussain SA, Billingham LJ, Murray PG, Deshmukh N, James ND,  
461 Wallace DM, Johnson PJ, Zeegers MP, Cheng KK, Martin A, Ward DG. Assessment of high-throughput high-  
462 resolution MALDI-TOF-MS of urinary peptides for the detection of muscle-invasive bladder cancer.  
463 *Proteomics Clin Appl* 2011; 5:493-503.

464 (51) Shimwell NJ, Bryan RT, Wei W, James ND, Cheng KK, Zeegers MP, Johnson PJ, Martin A, Ward DG. Combined  
465 proteome and transcriptome analyses for the discovery of urinary biomarkers for urothelial carcinoma. *Br J*  
466 *Cancer* 2013; 108:1854-1861.

467 (52) Bryan RT, Shimwell NJ, Wei W, Devall AJ, Pirrie SJ, James ND, Zeegers MP, Cheng KK, Martin A, Ward DG.  
468 Urinary EpCAM in urothelial bladder cancer patients: characterisation and evaluation of biomarker potential.  
469 *Br J Cancer* 2013.

470 (53) Hawkins RD, Hon GC, Ren B. Next-generation genomics: an integrative approach. *Nat Rev Genet* 2010;  
471 11:476-486.

472 (54) Svatek RS, Hollenbeck BK, Holmang S, Lee R, Kim SP, Stenzl A, Lotan Y. The Economics of Bladder Cancer:  
473 Costs and Considerations of Caring for This Disease. *Eur Urol* 2014.

474 (55) Choi W, Porten S, Kim S, Willis D, Plimack ER, Hoffman-Censits J, Roth B, Cheng T, Tran M, Lee IL, Melquist J,  
475 Bondaruk J, Majewski T, Zhang S, Pretzsch S, Baggerly K, Siefker-Radtke A, Czerniak B, Dinney CP, McConkey  
476 DJ. Identification of distinct basal and luminal subtypes of muscle-invasive bladder cancer with different  
477 sensitivities to frontline chemotherapy. *Cancer Cell* 2014; 25:152-165.

478 (56) Hurst CD, Knowles MA. Molecular subtyping of invasive bladder cancer: time to divide and rule? *Cancer Cell*  
479 2014; 25:135-136.

480 (57) Boustead GB, Fowler S, Swamy R, Kocklebergh R, Hounsborne L. Stage, grade and pathological characteristics  
481 of bladder cancer in the UK: British Association of Urological Surgeons (BAUS) Urological Tumour Registry.  
482 *BJU Int* 2013.

483 (58) Bryan RT, Zeegers MP, van Roekel EH, Bird D, Grant MR, Dunn JA, Bathers S, Iqbal G, Khan HS, Collins SI,  
484 Howman A, Deshmukh NS, James ND, Cheng KK, Wallace DM. A comparison of patient and tumour  
485 characteristics in two UK bladder cancer cohorts separated by 20 years. *BJU Int* 2013.

486 (59) Mao Q, Li Y, Zheng X, Yang K, Shen H, Qin J, Bai Y, Kong D, Jia X, Xie L. Up-regulation of E-cadherin by small  
487 activating RNA inhibits cell invasion and migration in 5637 human bladder cancer cells. *Biochem Biophys Res*  
488 *Commun* 2008; 375:566-570.

489 (60) Mialhe A, Levacher G, Champelovier P, Martel V, Serres M, Knudsen K, Seigneurin D. Expression of E-, P-, n-  
490 cadherins and catenins in human bladder carcinoma cell lines. *J Urol* 2000; 164:826-835.

491 (61) Wheelock MJ, Shintani Y, Maeda M, Fukumoto Y, Johnson KR. Cadherin switching. *J Cell Sci* 2008; 121:727-  
492 735.

493 (62) Nollet F, Kools P, van RF. Phylogenetic analysis of the cadherin superfamily allows identification of six major  
494 subfamilies besides several solitary members. *J Mol Biol* 2000; 299:551-572.

495 (63) Wheelock MJ, Knudsen KA. Cadherins and associated proteins. *In Vivo* 1991; 5:505-513.

496 (64) Brasch J, Harrison OJ, Honig B, Shapiro L. Thinking outside the cell: how cadherins drive adhesion. *Trends Cell*  
497 *Biol* 2012; 22:299-310.

498 (65) Fichtner D, Lorenz B, Engin S, Deichmann C, Oelkers M, Janshoff A, Menke A, Wedlich D, Franz CM. Covalent  
499 and density-controlled surface immobilization of e-cadherin for adhesion force spectroscopy. *PLoS One*  
500 2014; 9:e93123.

501 (66) Ismail AF, Dasgupta P, Khan MS, Jiang W, Martin T, Wells CM. P21-activated kinase 5 (PAK5) and epithelial  
502 mesenchymal transition in bladder cancer. *Eur Urol* 14 A.D..

503 (67) Zaidel-Bar R. Cadherin adhesome at a glance. *J Cell Sci* 2013; 126:373-378.

504 (68) Harrison OJ, Jin X, Hong S, Bahna F, Ahlsen G, Brasch J, Wu Y, Vendome J, Felsovalyi K, Hampton CM,  
505 Troyanovsky RB, Ben-Shaul A, Frank J, Troyanovsky SM, Shapiro L, Honig B. The extracellular architecture of  
506 adherens junctions revealed by crystal structures of type I cadherins. *Structure* 2011; 19:244-256.

507 (69) Rieger-Christ KM, Cain JW, Braasch JW, Dugan JM, Silverman ML, Bouyounes B, Libertino JA, Summerhayes  
508 IC. Expression of classic cadherins type I in urothelial neoplastic progression. *Hum Pathol* 2001; 32:18-23.

509 (70) Takeichi M. Cadherin cell adhesion receptors as a morphogenetic regulator. *Science* 1991; 251:1451-1455.

510 (71) Shimoyama Y, Hirohashi S, Hirano S, Noguchi M, Shimosato Y, Takeichi M, Abe O. Cadherin cell-adhesion  
511 molecules in human epithelial tissues and carcinomas. *Cancer Res* 1989; 49:2128-2133.

512 (72) Shiozaki H, Tahara H, Oka H, Miyata M, Kobayashi K, Tamura S, Iihara K, Doki Y, Hirano S, Takeichi M, .  
513 Expression of immunoreactive E-cadherin adhesion molecules in human cancers. *Am J Pathol* 1991; 139:17-  
514 23.

515 (73) Takeichi M. The cadherins: cell-cell adhesion molecules controlling animal morphogenesis. *Development*  
516 1988; 102:639-655.

517 (74) Paredes J, Figueiredo J, Albergaria A, Oliveira P, Carvalho J, Ribeiro AS, Caldeira J, Costa AM, Simoes-Correia  
518 J, Oliveira MJ, Pinheiro H, Pinho SS, Mateus R, Reis CA, Leite M, Fernandes MS, Schmitt F, Carneiro F,  
519 Figueiredo C, Oliveira C, Seruca R. Epithelial E- and P-cadherins: role and clinical significance in cancer.  
520 *Biochim Biophys Acta* 2012; 1826:297-311.

521 (75) van RF. Beyond E-cadherin: roles of other cadherin superfamily members in cancer. *Nat Rev Cancer* 2014;  
522 14:121-134.

523 (76) Usui A, Ko SY, Barengo N, Naora H. P-cadherin promotes ovarian cancer dissemination through tumor cell  
524 aggregation and tumor-peritoneum interactions. *Mol Cancer Res* 2014; 12:504-513.

525 (77) De WO, Pauwels P, De CB, Sabbah M, Emami S, Redeuilh G, Gespach C, Bracke M, Berx G. Molecular and  
526 pathological signatures of epithelial-mesenchymal transitions at the cancer invasion front. *Histochem Cell*  
527 *Biol* 2008; 130:481-494.

528 (78) Friedl P, Gilmour D. Collective cell migration in morphogenesis, regeneration and cancer. *Nat Rev Mol Cell*  
529 *Biol* 2009; 10:445-457.

530 (79) Chiang AC, Massague J. Molecular basis of metastasis. *N Engl J Med* 2008; 359:2814-2823.

531 (80) Tselepis C, Perry I, Jankowski J. Barrett's esophagus: dysregulation of cell cycling and intercellular adhesion in  
532 the metaplasia-dysplasia-carcinoma sequence. *Digestion* 2000; 61:1-5.

533 (81) Maeda M, Johnson KR, Wheelock MJ. Cadherin switching: essential for behavioral but not morphological  
534 changes during an epithelium-to-mesenchyme transition. *J Cell Sci* 2005; 118:873-887.

535 (82) Farahani E, Patra HK, Jangamreddy JR, Rashedi I, Kawalec M, Rao Pariti RK, Batakis P, Wiechec E. Cell  
536 adhesion molecules and their relation to (cancer) cell stemness. *Carcinogenesis* 2014; 35:747-759.

537 (83) Taniuchi K, Nakagawa H, Hosokawa M, Nakamura T, Eguchi H, Ohigashi H, Ishikawa O, Katagiri T, Nakamura  
538 Y. Overexpressed P-cadherin/CDH3 promotes motility of pancreatic cancer cells by interacting with p120ctn  
539 and activating rho-family GTPases. *Cancer Res* 2005; 65:3092-3099.

540 (84) Gravdal K, Halvorsen OJ, Haukaas SA, Akslen LA. A switch from E-cadherin to N-cadherin expression indicates  
541 epithelial to mesenchymal transition and is of strong and independent importance for the progress of  
542 prostate cancer. *Clin Cancer Res* 2007; 13:7003-7011.

543 (85) Jaggi M, Nazemi T, Abrahams NA, Baker JJ, Galich A, Smith LM, Balaji KC. N-cadherin switching occurs in high  
544 Gleason grade prostate cancer. *Prostate* 2006; 66:193-199.

545 (86) Patel IS, Madan P, Getsios S, Bertrand MA, MacCalman CD. Cadherin switching in ovarian cancer progression.  
546 *Int J Cancer* 2003; 106:172-177.

547 (87) Li G, Satyamoorthy K, Herlyn M. N-cadherin-mediated intercellular interactions promote survival and  
548 migration of melanoma cells. *Cancer Res* 2001; 61:3819-3825.

549 (88) Shimoyama Y, Hirohashi S, Hirano S, Noguchi M, Shimosato Y, Takeichi M, Abe O. Cadherin cell-adhesion  
550 molecules in human epithelial tissues and carcinomas. *Cancer Res* 1989; 49:2128-2133.

551 (89) Wang P, Lin SL, Zhang LH, Li Z, Liu Q, Gao JX, Liu DM, Bo JJ, Huang YR. The prognostic value of P-cadherin in  
552 non-muscle-invasive bladder cancer. *Eur J Surg Oncol* 2014; 40:255-259.

553 (90) Wu K, Zeng J, Zhou J, Fan J, Chen Y, Wang Z, Zhang T, Wang X, He D. Slug contributes to cadherin switch and  
554 malignant progression in muscle-invasive bladder cancer development. *Urol Oncol* 2013; 31:1751-1760.

555 (91) Lascombe I, Clairotte A, Fauconnet S, Bernardini S, Wallerand H, Kantelip B, Bittard H. N-cadherin as a novel  
556 prognostic marker of progression in superficial urothelial tumors. *Clin Cancer Res* 2006; 12:2780-2787.

557 (92) Mandeville JA, Silva NB, Vanni AJ, Smith GL, Rieger-Christ KM, Zeheb R, Loda M, Libertino JA, Summerhayes  
558 IC. P-cadherin as a prognostic indicator and a modulator of migratory behaviour in bladder carcinoma cells.  
559 *BJU Int* 2008; 102:1707-1714.

560 (93) Van BJ, Yamany T, Molotkov A, Dan H, Mansukhani M, Batourina E, Schneider K, Oyon D, Dunlop M, Wu XR,  
561 Cordon-Cardo C, Mendelsohn C. Bladder cancers arise from distinct urothelial sub-populations. *Nat Cell Biol*  
562 2014; 16:982-991.

563 (94) Sarrio D, Palacios J, Hergueta-Redondo M, Gomez-Lopez G, Cano A, Moreno-Bueno G. Functional  
564 characterization of E- and P-cadherin in invasive breast cancer cells. *BMC Cancer* 2009; 9:74.

565 (95) Ribeiro AS, Sousa B, Carreto L, Mendes N, Nobre AR, Ricardo S, Albergaria A, Cameselle-Teijeiro JF, Gerhard  
566 R, Soderberg O, Seruca R, Santos MA, Schmitt F, Paredes J. P-cadherin functional role is dependent on E-  
567 cadherin cellular context: a proof of concept using the breast cancer model. *J Pathol* 2013; 229:705-718.

568 (96) Paredes J, Albergaria A, Oliveira JT, Jeronimo C, Milanezi F, Schmitt FC. P-cadherin overexpression is an  
569 indicator of clinical outcome in invasive breast carcinomas and is associated with CDH3 promoter  
570 hypomethylation. *Clin Cancer Res* 2005; 11:5869-5877.

571 (97) Sato N, Fukushima N, Maitra A, Matsubayashi H, Yeo CJ, Cameron JL, Hruban RH, Goggins M. Discovery of  
572 novel targets for aberrant methylation in pancreatic carcinoma using high-throughput microarrays. *Cancer*  
573 *Res* 2003; 63:3735-3742.

574 (98) Milicic A, Harrison LA, Goodlad RA, Hardy RG, Nicholson AM, Presz M, Sieber O, Santander S, Pringle JH,  
575 Mandir N, East P, Obszynska J, Sanders S, Piazuelo E, Shaw J, Harrison R, Tomlinson IP, McDonald SA, Wright  
576 NA, Jankowski JA. Ectopic expression of P-cadherin correlates with promoter hypomethylation early in  
577 colorectal carcinogenesis and enhanced intestinal crypt fission in vivo. *Cancer Res* 2008; 68:7760-7768.

578 (99) Ribeiro AS, Albergaria A, Sousa B, Correia AL, Bracke M, Seruca R, Schmitt FC, Paredes J. Extracellular  
579 cleavage and shedding of P-cadherin: a mechanism underlying the invasive behaviour of breast cancer cells.  
580 *Oncogene* 2010; 29:392-402.

581 (100) Paredes J, Correia AL, Ribeiro AS, Milanezi F, Cameselle-Teijeiro J, Schmitt FC. Breast carcinomas that co-  
582 express E- and P-cadherin are associated with p120-catenin cytoplasmic localisation and poor patient  
583 survival. *J Clin Pathol* 2008; 61:856-862.

584 (101) Taniuchi K, Nakagawa H, Hosokawa M, Nakamura T, Eguchi H, Ohigashi H, Ishikawa O, Katagiri T, Nakamura  
585 Y. Overexpressed P-cadherin/CDH3 promotes motility of pancreatic cancer cells by interacting with p120ctn  
586 and activating rho-family GTPases. *Cancer Res* 2005; 65:3092-3099.

587 (102) Cheung LW, Leung PC, Wong AS. Cadherin switching and activation of p120 catenin signaling are mediators  
588 of gonadotropin-releasing hormone to promote tumor cell migration and invasion in ovarian cancer.  
589 *Oncogene* 2010; 29:2427-2440.

590 (103) Cheung LW, Mak AS, Cheung AN, Ngan HY, Leung PC, Wong AS. P-cadherin cooperates with insulin-like  
591 growth factor-1 receptor to promote metastatic signaling of gonadotropin-releasing hormone in ovarian  
592 cancer via p120 catenin. *Oncogene* 2011; 30:2964-2974.

593 (104) Zhang CC, Yan Z, Zhang Q, Kuszpit K, Zasadny K, Qiu M, Painter CL, Wong A, Kraynov E, Arango ME, Mehta  
594 PP, Popoff I, Casperson GF, Los G, Bender S, Anderes K, Christensen JG, VanArsdale T. PF-03732010: a fully  
595 human monoclonal antibody against P-cadherin with antitumor and antimetastatic activity. *Clin Cancer Res*  
596 2010; 16:5177-5188.

597 (105) Moltzahn FR, Volkmer JP, Rottke D, Ackermann R. "Cancer stem cells"-lessons from Hercules to fight the  
598 Hydra. *Urol Oncol* 2008; 26:581-589.

599 (106) Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature* 2001;  
600 414:105-111.

601 (107) Gupta PB, Chaffer CL, Weinberg RA. Cancer stem cells: mirage or reality? *Nature Medicine* 2009; 15:1010-  
602 1012.

603 (108) Brandt WD, Matsui W, Rosenberg JE, He X, Ling S, Schaeffer EM, Berman DM. Urothelial carcinoma: stem  
604 cells on the edge. *Cancer Metastasis Rev* 2009; 28:291-304.

605 (109) Ben-Porath I, Thomson MW, Carey VJ, Ge R, Bell GW, Regev A, Weinberg RA. An embryonic stem cell-like  
606 gene expression signature in poorly differentiated aggressive human tumors. *Nat Genet* 2008; 40:499-507.

607 (110) Visvader JE, Lindeman GJ. Cancer stem cells in solid tumours: accumulating evidence and unresolved  
608 questions. *Nat Rev Cancer* 2008; 8:755-768.

609 (111) Hoglund M. On the origin of syn- and metachronous urothelial carcinomas. *Eur Urol* 2007; 51:1185-1193.

610 (112) Chan KS, Espinosa I, Chao M, Wong D, Ailles L, Diehn M, Gill H, Presti J, Jr., Chang HY, van de RM, Shortliffe L,  
611 Weissman IL. Identification, molecular characterization, clinical prognosis, and therapeutic targeting of  
612 human bladder tumor-initiating cells. *Proc Natl Acad Sci U S A* 2009; 106:14016-14021.

613 (113) Rieger-Christ KM, Cain JW, Braasch JW, Dugan JM, Silverman ML, Bouyounes B, Libertino JA, Summerhayes  
614 IC. Expression of classic cadherins type I in urothelial neoplastic progression. *Hum Pathol* 2001; 32:18-23.

615 (114) Kolle G, Ho M, Zhou Q, Chy HS, Krishnan K, Cloonan N, Bertoncello I, Laslett AL, Grimmond SM. Identification  
616 of human embryonic stem cell surface markers by combined membrane-polysome translation state array  
617 analysis and immunotranscriptional profiling. *Stem Cells* 2009; 27:2446-2456.

618 (115) Vieira AF, Ricardo S, Ablett MP, Dionisio MR, Mendes N, Albergaria A, Farnie G, Gerhard R, Cameselle-  
619 Teijeiro JF, Seruca R, Schmitt F, Clarke RB, Paredes J. P-cadherin is coexpressed with CD44 and CD49f and  
620 mediates stem cell properties in basal-like breast cancer. *Stem Cells* 2012; 30:854-864.

621 (116) Bryan RT, Zeegers MP, James ND, Wallace DM, Cheng KK. Biomarkers in bladder cancer. *BJU Int* 2009.

622 (117) Rosen JM, Jordan CT. The increasing complexity of the cancer stem cell paradigm. *Science* 2009; 324:1670-  
623 1673.

624 (118) Szala S, Froehlich M, Scollon M, Kasai Y, Steplewski Z, Koprowski H, Linnenbach AJ. Molecular cloning of  
625 cDNA for the carcinoma-associated antigen GA733-2. *Proc Natl Acad Sci U S A* 1990; 87:3542-3546.

626 (119) Patriarca C, Colombo P, Pio TA, Wesseling J, Franchi G, Guddo F, Naspro R, Macchi RM, Giunta P, Di PM,  
627 Parente M, Arizzi C, Roncalli M, Campo B. Cell discohesion and multifocality of carcinoma in situ of the

628 bladder: new insight from the adhesion molecule profile (e-cadherin, Ep-CAM, and MUC1). *Int J Surg Pathol* 2009; 17:99-106.  
629

630 (120) Brunner A, Prelog M, Verdorfer I, Tzankov A, Mikuz G, Ensinger C. EpCAM is predominantly expressed in high  
631 grade and advanced stage urothelial carcinoma of the bladder. *J Clin Pathol* 2008; 61:307-310.

632 (121) Okegawa T, Hayashi K, Hara H, Nutahara K, Higashihara E. Immunomagnetic quantification of circulating  
633 tumor cells in patients with urothelial cancer. *Int J Urol* 2010; 17:254-258.

634 (122) Kowalski M, Entwistle J, Cizeau J, Niforos D, Loewen S, Chapman W, MacDonald GC. A Phase I study of an  
635 intravesically administered immunotoxin targeting EpCAM for the treatment of nonmuscle-invasive bladder  
636 cancer in BCGrefractory and BCG-intolerant patients. *Drug Des Devel Ther* 2010; 4:313-320.

637 (123) Maetzel D, Denzel S, Mack B, Canis M, Went P, Benk M, Kieu C, Papior P, Baeuerle PA, Munz M, Gires O.  
638 Nuclear signalling by tumour-associated antigen EpCAM. *Nat Cell Biol* 2009; 11:162-171.

639 (124) Chaves-Perez A, Mack B, Maetzel D, Kremling H, Eggert C, Harreus U, Gires O. EpCAM regulates cell cycle  
640 progression via control of cyclin D1 expression. *Oncogene* 2013; 32:641-650.

641 (125) Zeegers MP, Bryan RT, Langford C, Billingham L, Murray P, Deshmukh NS, Hussain S, James N, Wallace DM,  
642 Cheng KK. The West Midlands Bladder Cancer Prognosis Programme: rationale and design. *BJU Int* 2010;  
643 105:784-788.

644 (126) Ralhan R, He HC, So AK, Tripathi SC, Kumar M, Hasan MR, Kaur J, Kashat L, MacMillan C, Chauhan SS,  
645 Freeman JL, Walfish PG. Nuclear and cytoplasmic accumulation of Ep-ICD is frequently detected in human  
646 epithelial cancers. *PLoS One* 2010; 5:e14130.

647 (127) Cogliervina M, Guarnaccia C, Zlatev V, Pongor S, Pintar A. Jagged-1 juxtamembrane region: biochemical  
648 characterization and cleavage by ADAM17 (TACE) catalytic domain. *Biochem Biophys Res Commun* 2013;  
649 432:666-671.

650 (128) van der Gun BT, Melchers LJ, Ruiters MH, de Leij LF, McLaughlin PM, Rots MG. EpCAM in carcinogenesis: the  
651 good, the bad or the ugly. *Carcinogenesis* 2010; 31:1913-1921.

652 (129) Litvinov SV, Balzar M, Winter MJ, Bakker HA, Briaire-de Bruijn IH, Prins F, Fleuren GJ, Warnaar SO. Epithelial  
653 cell adhesion molecule (Ep-CAM) modulates cell-cell interactions mediated by classic cadherins. *J Cell Biol*  
654 1997; 139:1337-1348.

655 (130) Winter MJ, Nagelkerken B, Mertens AE, Rees-Bakker HA, Briaire-de Bruijn IH, Litvinov SV. Expression of Ep-  
656 CAM shifts the state of cadherin-mediated adhesions from strong to weak. *Exp Cell Res* 2003; 285:50-58.

657 (131) Winter MJ, Cirulli V, Briaire-de Bruijn IH, Litvinov SV. Cadherins are regulated by Ep-CAM via  
658 phosphatidylinositol-3 kinase. *Mol Cell Biochem* 2007; 302:19-26.

659 (132) Martowicz A, Spizzo G, Gastl G, Untergasser G. Phenotype-dependent effects of EpCAM expression on  
660 growth and invasion of human breast cancer cell lines. *BMC Cancer* 2012; 12:501.

661 (133) Martowicz A, Rainer J, Lelong J, Spizzo G, Gastl G, Untergasser G. EpCAM overexpression prolongs  
662 proliferative capacity of primary human breast epithelial cells and supports hyperplastic growth. *Mol Cancer*  
663 2013; 12:56.

664 (134) Slanchev K, Carney TJ, Stemmler MP, Koschorz B, Amsterdam A, Schwarz H, Hammerschmidt M. The  
665 epithelial cell adhesion molecule EpCAM is required for epithelial morphogenesis and integrity during  
666 zebrafish epiboly and skin development. *PLoS Genet* 2009; 5:e1000563.



667 (135) Guerra E, Lattanzio R, La SR, Dini F, Tiboni GM, Piantelli M, Alberti S. mTrop1/Epcam knockout mice develop  
668 congenital tufting enteropathy through dysregulation of intestinal E-cadherin/beta-catenin. PLoS One 2012;  
669 7:e49302.

670 (136) Maghzal N, Kayali HA, Rohani N, Kajava AV, Fagotto F. EpCAM controls actomyosin contractility and cell  
671 adhesion by direct inhibition of PKC. Dev Cell 2013; 27:263-277.

672 (137) Vannier C, Mock K, Brabletz T, Driever W. Zeb1 regulates E-cadherin and Epcam (epithelial cell adhesion  
673 molecule) expression to control cell behavior in early zebrafish development. J Biol Chem 2013; 288:18643-  
674 18659.

675 (138) Huang HP, Chen PH, Yu CY, Chuang CY, Stone L, Hsiao WC, Li CL, Tsai SC, Chen KY, Chen HF, Ho HN, Kuo HC.  
676 Epithelial cell adhesion molecule (EpCAM) complex proteins promote transcription factor-mediated  
677 pluripotency reprogramming. J Biol Chem 2011; 286:33520-33532.

678 (139) Chen HF, Chuang CY, Lee WC, Huang HP, Wu HC, Ho HN, Chen YJ, Kuo HC. Surface marker epithelial cell  
679 adhesion molecule and E-cadherin facilitate the identification and selection of induced pluripotent stem  
680 cells. Stem Cell Rev 2011; 7:722-735.

681 (140) Ravindran G, Sawant SS, Hague A, Kingsley K, Devaraj H. Association of differential beta-catenin expression  
682 with Oct-4 and Nanog in Oral Squamous Cell Carcinoma and their correlation with Clinicopathological factors  
683 and Prognosis. Head Neck 2014.

684 (141) Luo Y, Lan L, Jiang YG, Zhao JH, Li MC, Wei NB, Lin YH. Epithelial-mesenchymal transition and migration of  
685 prostate cancer stem cells is driven by cancer-associated fibroblasts in an HIF-1alpha/beta-catenin-  
686 dependent pathway. Mol Cells 2013; 36:138-144.

687 (142) He K, Xu T, Xu Y, Ring A, Kahn M, Goldkorn A. Cancer cells acquire a drug resistant, highly tumorigenic,  
688 cancer stem-like phenotype through modulation of the PI3K/Akt/beta-catenin/CBP pathway. Int J Cancer  
689 2014; 134:43-54.

690 (143) Macaulay IC, Voet T. Single cell genomics: advances and future perspectives. PLoS Genet 2014; 10:e1004126.

691 (144) Van LP, Voet T. Single cell analysis of cancer genomes. Curr Opin Genet Dev 2014; 24C:82-91.

692 (145) Vogelstein B, Papadopoulos N, Velculescu VE, Zhou S, Diaz LA, Jr., Kinzler KW. Cancer genome landscapes.  
693 Science 2013; 339:1546-1558.

694 (146) Blaschuk OW. Discovery and development of N-cadherin antagonists. Cell Tissue Res 2012; 348:309-313.

695 (147) Devemy E, Blaschuk OW. Identification of a novel N-cadherin antagonist. Peptides 2008; 29:1853-1861.

696 (148) Blaschuk OW, Devemy E. Cadherins as novel targets for anti-cancer therapy. Eur J Pharmacol 2009; 625:195-  
697 198.

698 (149) Warneke VS, Behrens HM, Haag J, Kruger S, Simon E, Mathiak M, Ebert MP, Rocken C. Members of the  
699 EpCAM signalling pathway are expressed in gastric cancer tissue and are correlated with patient prognosis.  
700 Br J Cancer 2013; 109:2217-2227.

701 (150) Kleiber K, Strebhardt K, Martin BT. The biological relevance of FHL2 in tumour cells and its role as a putative  
702 cancer target. Anticancer Res 2007; 27:55-61.

703

704

705

## LEGENDS FOR FIGURES

**Figure 1: Cell-cell adhesion in epithelial tissues** (taken from [22]). **a)** overview of cell-cell adhesion complexes; **b)** pictorial representation of cell-cell interactions on neighbouring cells; **c)** molecular structure of the adherens junction, showing the relationship between E-cadherin molecules on neighbouring cells, and between E-cadherin, the catenins ( $\alpha$ ,  $\beta$ ,  $\gamma$ , p120) and the cell cytoskeleton. Traditionally, cadherins on neighbouring cells adhere via EC1 domains, although more recent research suggests that all 5 EC domains are required for optimal adhesion [65].

**Figure 2: Cadherin switching in bladder UCs** (taken from [22]). **a)** E-cadherin is strongly expressed at the cell membrane throughout the normal urothelium. Reduced expression is observed in a proportion of NMIBCs, and the majority of MIBCs demonstrate either reduced expression or a complete absence of E-cadherin; **b)** P-cadherin is expressed in the basal 1-2 layers of normal urothelium, and this pattern is preserved in the majority of NMIBCs. The majority of MIBCs demonstrate strong P-cadherin expression throughout the tumour mass; **c)** N-cadherin is not expressed in normal urothelium or the majority of NMIBCs. However, the majority of muscle-invasive UCs express N-cadherin throughout the tumour mass.

**Figure 3: Proposed pathways for the development of a bladder cancer stem cell phenotype and the relationship with EpCAM** (adapted from [23]). Cancer stem cells (CSCs) result in the development of treatment resistant disease in some cancer settings, and this diagram proposes potential pathways for their development in UBC. There is likely considerable plasticity in these pathways [142], with cells reverting to a less aggressive state by mesenchymal-to-epithelial transition (MET) or by the reversal of the CSC phenotype, and most likely influenced by the tumour microenvironment [23]. We also propose a model whereby EpCAM modulates the development of EMT and/or CSCs (see text).

**Figure 4: EpCAM's relationship with E-cadherin** (adapted from [123;149]). The dual role of EPCAM in epithelial tissues is demonstrated. EpCAM can either disrupt the adherens junction, resulting in the release of  $\beta$ -catenin **(a)**, or stabilise the adherens junction to maintain E-cadherin's anchorage to the cell cytoskeleton **(b)**. In **(a)**, released  $\beta$ -catenin subsequently forms a complex with EpiCD and the transcriptional co-factor FHL2 [150], either at the cell membrane or in the cell nucleus. The EpiCD/FHL2/ $\beta$ -catenin complex then interacts with the Lef-1 transcription

731 factor in the cell nucleus to activate the transcription of various target genes, including known oncogenes. In UBC we  
732 demonstrated that the extracellular domain of EpCAM is released by cleavage immediately adjacent to the cell  
733 membrane [52]. The exact location of cleavage was not described by Maetzel et al [123], but the protease involved  
734 (TACE or ADAM 17) usually cleaves membrane proteins 10-15 residues away from the membrane surface [127],  
735 suggesting atypical cleavage or an alternative mechanism of extracellular domain release in UBC. ( $\alpha$ = $\alpha$ -catenin,  $\beta$ = $\beta$ -  
736 catenin).

Figure a

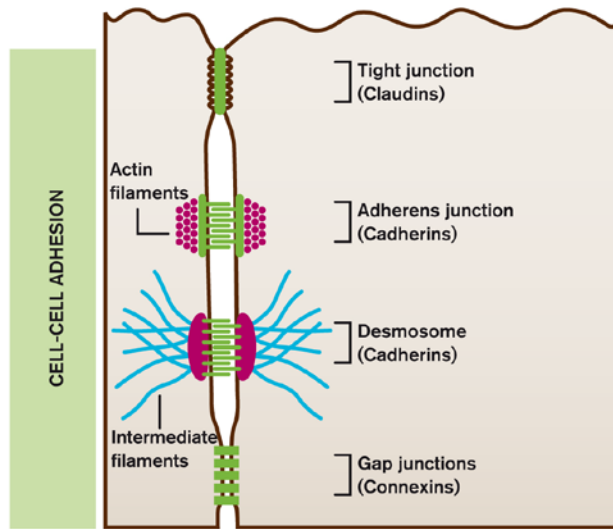


Figure b

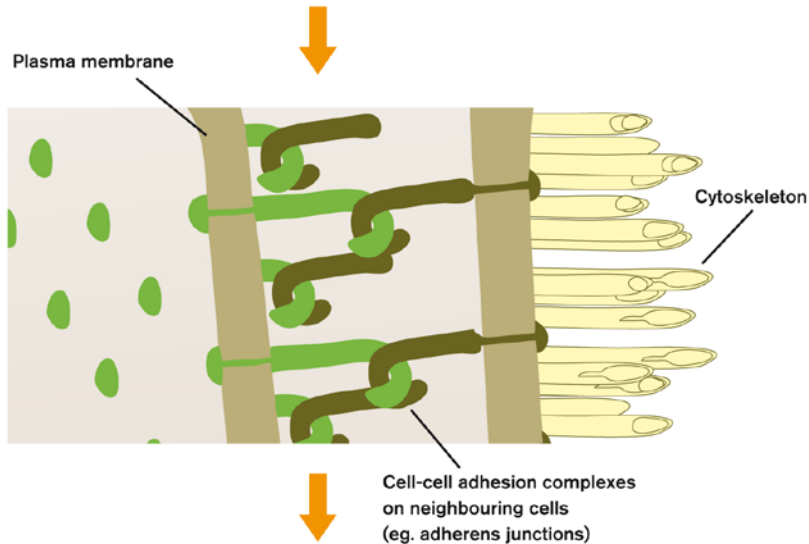


Figure c

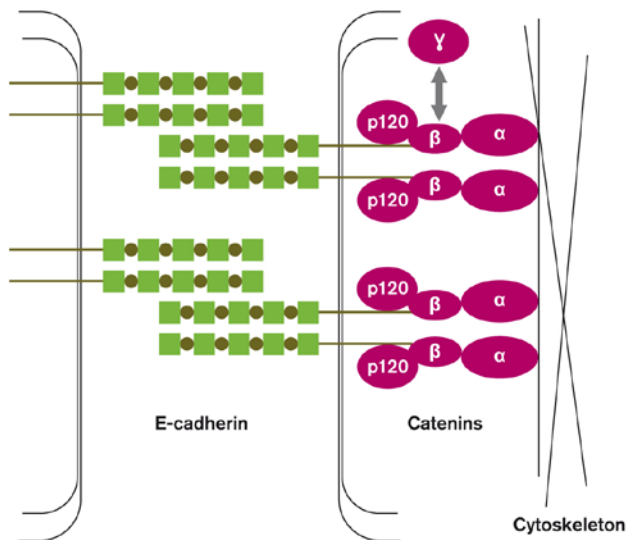
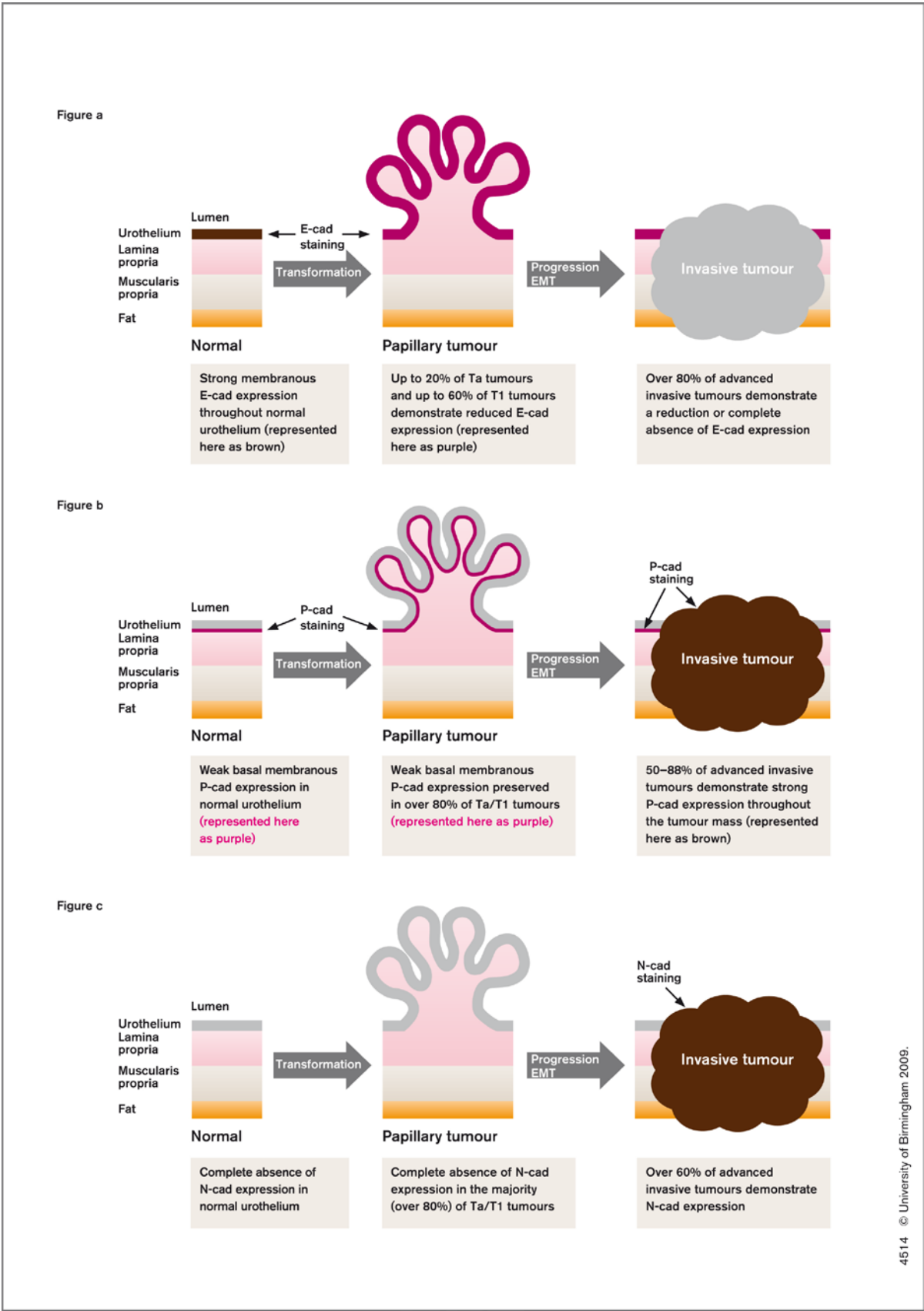
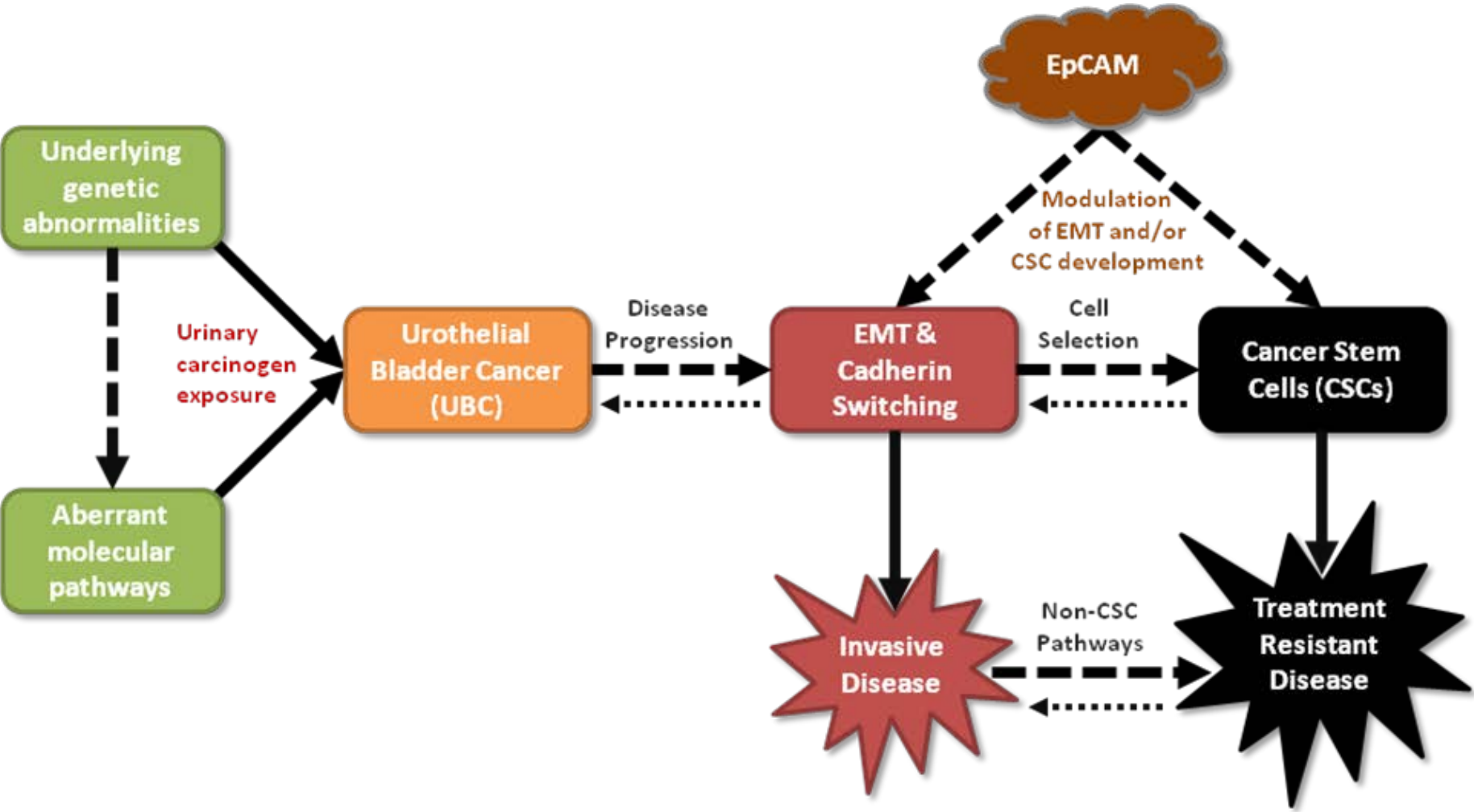


Figure 2: Cadherin switching in bladder UBCs.



744 Figure 3: Proposed pathways for the development of a bladder cancer stem cell phenotype and the relationship with EpCAM.

745



746

Figure 4: EpCAM's relationship with E-cadherin.

